The Paleozoic origin of enzymatic mechanisms for lignin degradation reconstructed using 31 fungal genomes

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The Paleozoic origin of enzymatic mechanisms for lignin degradation reconstructed using 31 fungal genomes

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Wood is a major pool of organic carbon that is highly resistant to decay, owing largely to the presence of lignin. The only organisms capable of significant lignin decay are white rot Fungi in the Agaricomycetes, which also contains non-lignin-degrading brown rot and ectomycorrhizal species. Comparative analyses of 31 fungal genomes (12 newly generated) suggest that lignin-degrading peroxidases expanded in the lineage leading to the ancestor of the Agaricomycetes, which is reconstructed as a white rot species, and then contracted in parallel lineages leading to brown rot and mycorrhizal species. Molecular clock analyses suggest that the origin of lignin degradation roughly coincided with the sharp decrease in the rate of organic carbon burial around the end of the Carboniferous period.

Lignin is a heterogeneous polymer that provides strength and rigidity to wood, protects cellulose and hemicellulose from microbial attack (*1*), and is the major precursor of coal (*2*). Genomic studies of wood decay organisms have focused on model systems for white rot (in which all plant cell wall components are degraded), such as *Phanerochaete chrysosporium (3),* and brown rot (in which lignin is modified but not appreciably degraded), such as *Postia placenta (4)* and *Serpula lacrymans (5)*. However, these species represent just two of the 18 recognized orders of Agaricomycetes, of which five contain brown rot taxa. To reconstruct the evolution of lignin decay mechanisms, we

analyzed 31 diverse fungal genomes, including twelve newly-sequenced species of Agaricomycotina (Table 1). The new genomes comprise six white rot species, five brown rot species, and one mycoparasite, representing nine orders (Figs. 1, S1-5; see Supporting Online Material for information on strains, culture conditions, genome size and assembly).

To estimate phylogenetic relationships, we constructed datasets using 71 or 26 single-copy genes, with varying alignment criteria and treatments for fast-evolving sites, yielding matrices of 10002 to 34257 amino acids, which we analyzed with maximum likelihood (ML) and Bayesian methods (*6, 7*) (see Supporting Online Material for details). All but six nodes receive maximal support values in all analyses and the rest are strongly supported (bootstrap ≥99% or posterior probability ≥0.99) in at least three analyses. The tree topology is consistent with prior analyses (*8*) and resolves four independent brown rot lineages (Figs. 1a, S6).

We next searched all 31 genomes for 27 gene families encoding oxidoreductases and carbohydrate-active enzymes (CAZymes) that have been implicated in wood decay (*9, 10*) (Table 1). CAZymes, particularly those acting on crystalline cellulose, are abundant in white rot genomes, which have 61-148 (avg. 87) copies of genes encoding CAZymes, representing 14-17 gene families, whereas brown rot genomes have 32-68 copies (avg. 46) from 9-12 families. The ectomycorrhizal *Laccaria bicolor* resembles brown rot species in this regard, possessing 28 CAZyme genes in eight families (Table 1). Notably, glycoside hydrolase (GH) families GH6 and GH7, which include cellobiohydrolases that are involved in the attack of crystalline cellulose (*9*), are present in all white rot lineages, but they are absent in brown rot

lineages (except Boletales) and *L. bicolor*. A similar pattern of enrichment in white rot genomes is shown by genes encoding GH61 enzymes, which have a copper-dependent oxidative mechanism for disrupting crystalline cellulose (*11*), and cellulose binding modules (CBM1), which effectively increase the concentration of the enzymes on the surface of crystalline cellulose (*12*) (Table 1).

Here, we focus on fungal class II peroxidases (PODs), which have been shown to be important in lignin degradation in *P. chrysosporium* and others (*13-15*). (The remaining gene families are discussed in Supporting Online Material; Figs. S7-19.) We classified PODs in four major groups, including three ligninolytic forms, lignin peroxidase (LiP), manganese peroxidase (MnP) and versatile peroxidase (VP), and a fourth POD type, defined here as “generic peroxidase” (GP), which is expected to include non-ligninolytic low redox-potential peroxidases with catalytic properties similar to those of the peroxidase of *Coprinopsis cinerea* or the product of the *nop*A gene in *P. chrysosporium* (*14*) (further subclassification of PODs is described in Supporting Online Material).

Consistent with a central role for PODs in lignin degradation, white rot species have 5 to 26 copies (avg. 14) of genes encoding ligninolytic PODs, but all brown rot species lack these enzymes, as do the ectomycorrhizal (ECM) *Laccaria bicolor,* the soil saprotroph *C. cinerea,* and *Schizophyllum commune,* which has been regarded as a white rot fungus, but has a limited capacity to degrade lignin (*15*). Moreover, analyses of gene diversification using binary state speciation analysis (*19*) confirmed that the rate of duplication of POD genes is elevated in white rot lineages vs. non-white rot lineages (see Supporting Online Material for details).

LiPs possess a tryptophan residue on the surface of the enzyme corresponding to Trp171 in *P. chrysosporium* LiP-H8 that enables direct oxidation of lignin compounds via long-range electron transfer, and MnPs possess two or three residues corresponding to Glu35, Glu39 and Asp175 of *P. chrysosporium* MnP1 that function in binding Mn (*16, 17*). VPs possess both the Trp171 homologue and Mn-binding residues, while all are lacking in GPs. To reconstruct functional evolution of PODs, we performed Bayesian and ML analyses (*6, 18*) using the GPs of Ascomycota as outgroups and reconstructed the evolution of the key residues using BayesTraits (*20*). The ancestor of all PODs lacked the Mn-binding and Trp171 residues, suggesting that it was non-ligninolytic (Fig. 1b). The most recent common ancestor of all ligninolytic Agaricomycete PODs is reconstructed as an MnP, with a single origin of LiP (gain of Trp171 and loss of Mn-binding residues), leading to parallel expansions in *P. chrysosporium* and *T. versicolor* (each with 10 LiP copies; Figs. 1b, S7, S17). Two origins of VP are resolved in the Polyporales, leading to *T. versicolor* and *D. squalens* (each with three VP copies; Figs. 1b, S7). VPs are also produced in the “oyster mushroom” *Pleurotus ostreatus* (Agaricales) (*21*), indicating further convergent evolution of this class of enzymes.

To localize the diversification of PODs in the organismal phylogeny, we performed gene tree/species tree reconciliation analyses using CAFÉ (*22*), Notung (*23*), and DrML (*24*). All methods suggest that a single POD gene copy was present in the common ancestor of Basidiomycota, with parallel losses in lineages leading to the Pucciniomycotina, Ustilaginomycotina, Tremellomycetes, and *Dacryopinax sp.* (Fig. 1a). Diversification of PODs began in the lineage leading to the most recent common

ancestor of the Agaricomycetes (node “A” in Fig. 1a), which is reconstructed as having two to seven POD gene copies in the various analyses. The “backbone” nodes in the Agaricomycete phylogeny (nodes labeled “B” in Fig. 1a) are reconstructed as having 3-16 POD gene copies, suggesting that duplicated PODs were retained throughout the early evolution of Agaricomycetes. Subsequently, all analyses suggest that there were parallel expansions of POD genes in the terminal lineages leading to at least seven of the eight white rot species, as well as parallel contractions within lineages leading to the brown rot species *Dacryopinax sp.*, *Gloeophyllum trabeum*, the Boletales, and the brown rot Polyporales (Fig. 1).

In addition to multiple PODs, reconciliation analyses suggest that the ancestor of the Agaricomycetes possessed one or two genes encoding dye-decolorizing peroxidases (DyP), which are heme peroxidases that have been shown to degrade lignin model compounds (*25, 26*), as well 5-8 genes encoding oxidases (including glyoxal oxidase) involved in peroxide generation (*27*) (details in Supporting Online Material). Collectively, these results suggest that the ancestor of Agaricomycetes was a white rot species that possessed a ligninolytic system with PODs, DyPs, and multiple pathways for H_2O_2 production.

To place the origin of lignin degradation in the context of geologic time, we performed Bayesian relaxed molecular clock analyses using BEAST (*18*) and PhyloBayes (*7*), with fossil-based calibrations at three nodes, including the ancestors of the Boletales (*28*), Agaricales (*29*), and Ascomycota (*30*) (see Supporting Online Material for details of molecular clock analyses). The mean age of the Agaricomycetes is ca. 290 mya in both BEAST and PhyloBayes analyses, with the mean age of the

Agaricomycotina being ca. 430-470 mya. These age estimates are consistent with basidiomycete fossils that were not used as calibration points, including hyphae with clamp connections from the Mississippian (*31*) (ca. 330 mya). BEAST analyses of the POD genes, calibrated with the split between Ascomycota and Basidiomycota according to the organismal phylogeny, suggest that the first ligninolytic MnP arose ca. 295 mya (Fig. 1), which is slightly earlier than (and therefore consistent with) the oldest definitive white rot fossils from the Permian (ca. 260 mya) and Triassic (ca. 230 mya) (*32*).

Organic carbon derived from lignin accumulated at an exceptionally high rate during the Carboniferous and Permian, resulting in the formation of vast coal deposits (*33*). A frequently cited explanation for this phenomenon is that decay was inhibited in the anoxic sediments of the widespread Permo-Carboniferous swamp forests. Our results are consistent with a complementary hypothesis (*2*), which posits that the sharp decline in the rate of organic carbon burial at the end of the Permo-Carboniferous was caused, at least in part, by the evolution of lignin decay capabilities in white rot Agaricomycetes.

Figure 1. **A.** Organismal phylogeny (chronogram) produced with BEAST. Light blue bars are 95% highest posterior density intervals for node ages; mean ages of selected nodes (millions of years) are in parentheses. Blue and red branches indicate significant expansion and contraction, respectively, of PODs inferred using CAFÉ. Numbers in red following following taxon names are POD gene counts. Numbers in red at nodes separated by commas, are numbers of POD gene copies estimated with CAFÉ, Notung (with two different edge weight threshold settings), and DrML, respectively. Node labled

A is the ancestor of Agaricomycetes, while nodes labeled B are "backbone" nodes in Agaricomycetes (see text). Asterisks indicate nodes that do not receive maximal support in all analyses (support values are in Fig. S6). Full species names are listed in a footnote to Table 1. **B.** POD gene phylogeny estimated in BEAST with ancestral state reconstructions for manganese-binding site (colored pies) and Trp171 residues (black and white pies) estimated with BayesTraits. Bars to right of gene IDs indicate presence of functional residues (*17*). Mean ages for selected nodes in parentheses are followed by 95% highest posterior density ranges.

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Supporting Online Material

Supplementary Methods, Results and Discussion

Tables S1 to S16

Figs. S1 to S22

References

Table 1. Gene contents in 17 CAZyme and 11 oxidoreductase families in the genomes of 20 Agaricomycotina and eleven other Fungi.

Taxonomy[1]		Basidiomycota																								Ascomycota					Ch	Mu
		Agaricomycotina																				Ust		Pucc		Pez				Sc		
		Agaricomycetes																	Dac	Trem												
		Ha	Sh	Pu	Fm	Ad	Tv	Ds	Pc	Pp	Wc	Fp	Gt	Sl	Cp	Lb	Sc	Cc	Da	Tm	Cn	Mg	Um	Ml	Sr	An	Cr	Tr	Sn	Ps	Bd	Pb
Ecology[2]		WR								BR						ECM	WR	S	BR	MP	AP	AP	PP	PP	Y	AP	PP	S	PP	Y	AP	S
CAZymes																																
Genes[3]	*p*[4]																															
GH3[7]	0.000	11	15	12	7	12	11	7	9	5	7	12	9	9	12	2	11	7	8	3	7	1	3	3	3	16	15	11	16	7	1	2
GH5[8]	0.305	7	6	6	6	8	5	5	5	5	4	5	5	8	8	3	3	6	5	0	0	0	1	7	2	4	7	3	5	0	1	1
GH6	0.916	1	1	1	2	2	1	1	1	0	0	0	0	1	2	0	1	5	0	0	0	0	0	0	0	2	2	1	4	0	0	0
GH7	0.000	1	3	5	2	6	4	4	8	0	0	0	0	0	2	0	2	6	0	0	0	0	0	9	0	2	5	2	5	0	0	0
GH10	0.004	2	6	5	4	4	6	5	6	4	4	2	3	1	3	0	5	6	3	0	0	0	2	6	0	2	4	1	7	1	0	0
GH11	0.687	0	1	1	0	3	0	0	1	0	0	0	0	0	0	0	1	6	0	0	0	0	1	0	0	3	4	3	7	0	0	0
GH12	0.483	4	5	2	3	1	5	3	2	2	2	2	2	1	4	3	1	1	1	0	0	0	0	5	0	3	5	2	4	0	0	0
GH28	0.000	8	17	13	16	10	11	7	4	7	9	13[9]	10	7	13	7	3	3	6	0	1	0	1	3	0	22	21	4	4	0	0	9
"GH"61	0.000	10	16	14	13	19	18	15	15	2	2	4	4	5	10	5	22	35	0	0	1	0	0	0	0	7	12	3	29	0	0	0
GH74	0.923	1	2	2	4	1	1	1	4	0	0	0	1	1	0	0	1	1	0	0	0	0	0	0	0	1	2	1	0	0	0	0
GH43	0.000	4	10	7	6	26	3	7	4	1[10]	1[10]	7	5	1	6	0	12	4	5	0	0	1	2	4	0	7	14	2	10	0	0	0
CE1	0.129	1	1	2	0	3	3	0	4	0	0	0	1	0	0	0	4	3	0	0	0	0	1	0	0	1	2	0	6	0	0	0
CE16	0.000	5	10	8	6	29	7	10	2	5	6	11	6	3	6	3	10	5	4	1	0	0	0	0	2	3	7	2	2	0	0	2
CE5	0.002	0	1	1	0	3	0	0	0	0	0	0	0	0	1	1	2	6	0	0	0	0	4	14	0	5	15	4	11	0	0	0
CE8	0.347	3	4	6	3	3	2	3	2	1	1	2	2	2	2	4	2	0	3	0	0	0	1	5	0	3	4	0	2	0	0	4
CE12	0.796	2	3	0	2	1	0	2	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	1	2	0	1	0	0	0
CE15	0.488	1	1	2	1	6	2	2	2	1	1	1	1	0	0	0	2	8	1	0	0	0	0	0	0	0	2	1	1	0	0	0
Oxidoreductases																																
POD	0.000	8	6	11	17	19	26	12	16	1	1	1	0	0	0	1	0	1	0	0	0	0	0	0	0	0	1	0	5	0	0	0

MCO	**0.000**	17	20	13	11	10	10	13	5	5	5	7	4	6	8	11	6	17	5	4	5	9	6	20	1	14	16	8	9	2	3	5
CRO[5]	**0.000**	5	8	9	4	9[4]	9	9	7	3	4	4	2	3	6	11	2	6	3	2	3	1	3	4	3	0	0	1	2	0	3	1
CDH[6]	**0.575**	1	1	1	1	1	1	1	1	0	0	0	1	2	2	0	1	1	0	0	0	0	0	0	0	2	3	0	3	0	0	0
Cytb562	**0.177**	1	1	0	0	0	1	1	1	0	0	0	0	2	3	0	2	0	0	0	0	0	0	0	0	0	1	0	2	0	0	0
OXO	**0.026**	3	3	2	3	3	5	5	7	5	4	5	4	3	2	1	5	1	2	0	0	0	0	1	0	2	2	3	3	0	0	0
GLP	**0.000**	1	11	6	8	1	2	6	3	5	10	10	6	3	11	1	7	0	3	0	0	0	0	2	0	0	0	0	0	0	0	0
QRD	**0.496**	2	1	3	3	4	1	1	4	1	1	1	3	2	2	2	4	3	1	1	2	0	1	1	1	1	1	1	1	4	1	4
DyP	**0.000**	1	2	5	3	11	2	1	0	2	0	0	0	0	0	2	0	4	0	1	1	0	0	2	0	0	0	0	0	0	0	0
HTP	**0.000**	5	10	8	4	16	3	4	3	5	5	4	6	3	2	5	3	8	6	0	0	0	3	17	0	5	2	4	13	0	0	0
P450	**0.000**	144	215	144	130	249	190	187	149	250	206	190	130	164	238	101	115	139	126	9	13	7	17	28	7	156	125	71	125	10	9	52

[1]Species: New genomes: Ad = *A. delicata*, Cp = *C. puteana*, Da = *Dacryopinax sp*., Ds = *D. squalens*, Fm = *F. mediterranea*, Fp = *F. pinicola*, Gt = *G. trabeum*, Pu = *P. strigosozonata*, Sh = *S. hirsutum*, Tm = *T. mesenterica*, Tv = *T. versicolor*, Wc = *W. cocos.* Others: An = *A. niger*, Bd = *B. dendrobatidis*, Cc = *C. cinerea*, Cn = *C. neoformans*, Cr = *C. parasitica*, Ha = *H. annosum* (has been reclassified as *H. irregulare*), Lb = *L. bicolor*, Mg = *M. globosa*, Ml = *M. laricis-populina*, Pb = *P. blakesleeanus*., Pc = *P. chrysosporium*, Pp = *P. placenta*, Ps = *P. stipitis*, Sc = *S. commune*, Sl = *S. lacrymans*, Sn = *S. nodorum*, Sr = *S. roseus*, Tr = *T. reesei*, Um = *U. maydis*. Higher taxa: Dac = Dacrymycetes, Trem = Tremellomycetes, Ust = Ustilaginomycotina, Pucc = Pucciniomycotina, Pez = Pezizomycotina, Sc = Saccharomycotina, Ch = Chytridiomycota, Mu = Mucoromycotina.

[2]WR = white rot, BR = brown rot, ECM = mycorrhiza, S = non-wood decay saprotroph, MP = mycoparasite, AP = animal pathogen/parasite, PP = plant pathogen, Y = yeast.

[3]POD= Clas-II peroxidases, MCO= multicopper oxidases, CRO= copper-radical oxidases, CDH= cellobiose dehydrogenase, Cytb562= cytochrome b562, OXO= oxalate oxidase/decarboxylases, GLP= Fe(III)-reducing glycopeptides, QRD= quinone reductases, DyP= dye-decolorizing peroxidases, HTP= heme-thiolate peroxidases, P450= cytochromes P450.

[4]*p*-values indicate strength of rejection of model of random diversification in CAFÉ analyses.

[5]One of four CRO2 genes in *A. delicata* is a potential pseudogene.

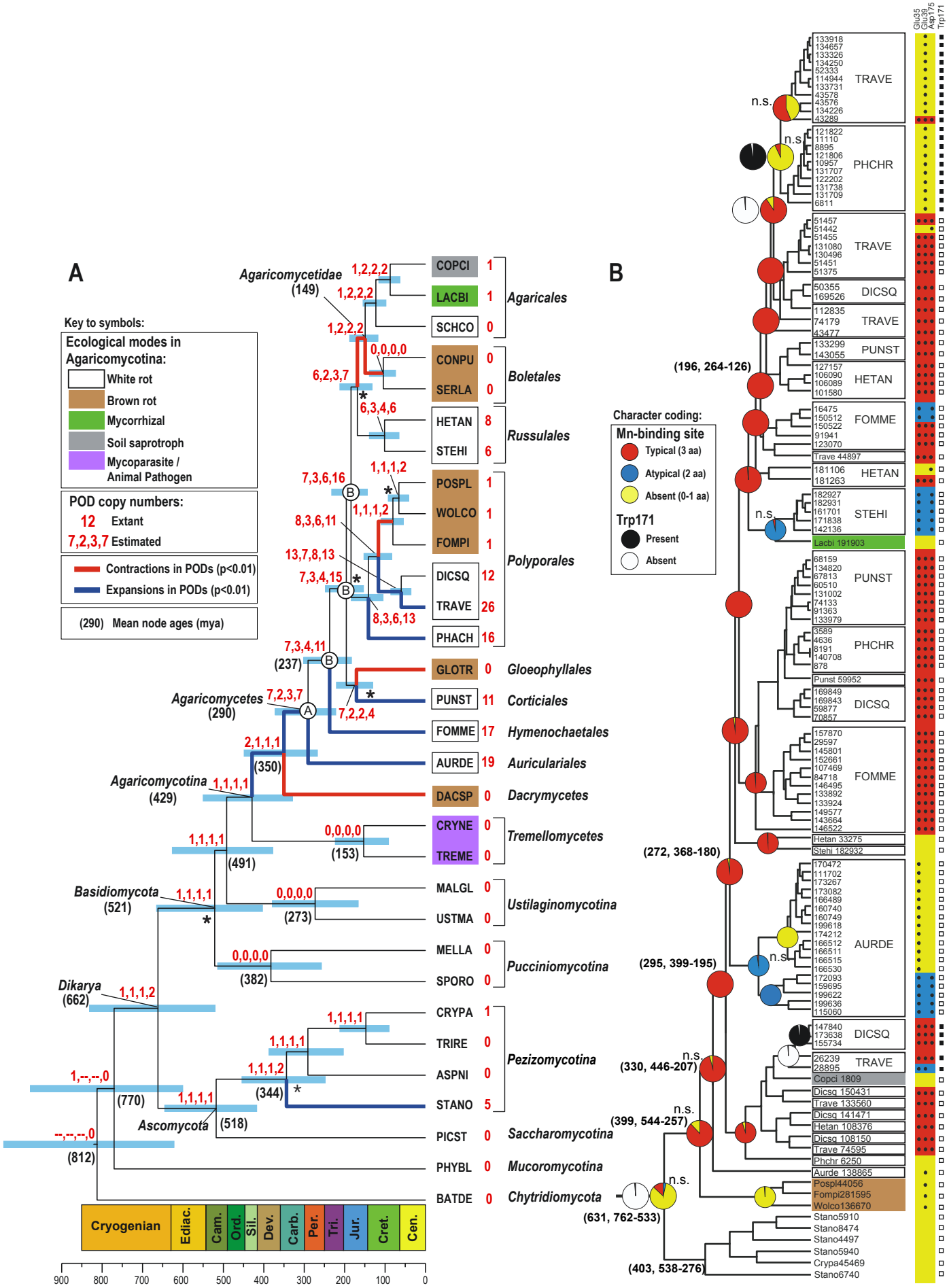
[6]One CDH gene in *C. puteana* lacks a cyt domain and may not be functional.

[7]GH3 does not include β-N-acetylhexosaminidase genes

[8]GH5 includes only models with similarity to endo-1,4-β-D-glucanases and mannan endo-β-1,4-mannosidases.

[9]One model (Fompi1 162677) is a potential pseudogene.

[10]The two models of Pospl1 and Wolco1 were missing from the original dataset as the clusters failed to recover them.


A
Key to symbols:
Ecological modes in Agaricomycotina:
White rot
Brown rot
Mycorrhizal
Soil saprotroph
Mycoparasite / Animal Pathogen
POD copy numbers:
12 Extant
7,2,3,7 Estimated
Contractions in PODs (p<0.01)
Expansions in PODs (p<0.01)
(290) Mean node ages (mya)
Agaricomycetidae (149)
Agaricomycetes (290)
Agaricomycotina (429)
Basidiomycota (521)
Dikarya (662)
Ascomycota (518)
COPCI 1
LACBI 1
SCHCO 0
Agaricales
CONPU 0
SERLA 0
Boletales
HETAN 8
STEHI 6
Russulales
POSPL 1
WOLCO 1
FOMPI 1
DICSQ 12
TRAVE 26
PHACH 16
Polyporales
GLOTR 0
Gloeophyllales
PUNST 11
Corticiales
FOMME 17
Hymenochaetales
AURDE 19
Auriculariales
DACSP 0
Dacrymycetes
CRYNE 0
TREME 0
Tremellomycetes
MALGL 0
USTMA 0
Ustilaginomycotina
MELLA 0
SPORO 0
Pucciniomycotina
CRYPA 1
TRIRE 0
ASPNI 0
STANO 5
Pezizomycotina
PICST 0
Saccharomycotina
PHYBL 0
Mucoromycotina
BATDE 0
Chytridiomycota
Cryogenian
Ediac.
Cam.
Ord.
Sil.
Dev.
Carb.
Per.
Tri.
Jur.
Cret.
Cen.
900 800 700 600 500 400 300 200 100 0
B
Character coding:
Mn-binding site
Typical (3 aa)
Atypical (2 aa)
Absent (0-1 aa)
Trp171
Present
Absent
Glu35
Glu39
Asp175
Trp171
TRAVE
PHCHR
DICSQ
PUNST
HETAN
FOMME
STEHI
AURDE
n.s.
(196, 264-126)
(272, 368-180)
(295, 399-195)
(330, 446-207)
(399, 544-257)
(631, 762-533)
(403, 538-276)
Copci 1809
Lacbi 191903
Pospi44056
Fompi281595
Wolco136670
Stano5910
Stano8474
Stano4497
Stano5940
Crypa45469
Stano6740